

**APPLICATION FOR AN EXEMPT DEALING**

Please attach Risk Assessments, plasmid maps and / or manuals pertinent to the GMO for which you are making application to the IBC.

**1. Project Title**

<Project Title>

**2. Project Supervisor**

<Name of Project Supervisor>

School: <Details of School and campus>  
 Building and Room No.: <Building and room No.>  
 Phone: <Work phone number>  
 Email: <work email address>

**3. Project Dates** (please note a project must not begin before the IBC has given approval)

Commencement Date: dd/mm/yy

Finishing Date: dd/mm/yy

**4. Exemption Category** (please mark applicable category with an x)

*\*refer to table at end of this form for the detailed description of each category.*

1-2	Dealing with a genetically modified <i>Caenorhabditis elegans</i>	
1-3	Dealing with an animal into which genetically modified somatic cells have been introduced	
1-3A	Dealing with an animal whose somatic cells have been genetically modified in vivo by a replication defective viral vector	
1-4	Dealing involving a host / vector system	
	HOST / VECTOR DETAILS Item: Class: Host: Vector:	
1-5	Dealing involving a shotgun cloning OR the preparation of a cDNA library, in a host / vector system	

**4.1 Modified trait(s) – class and details**

<response>.

**4.2 Details of recombinant DNA, including vector**

<response, including brief summary of procedures if relevant>.

**5. PC Level:**

**6. Facilities to be used**

Facility Name:

Building No.

Room No.

**7. Method of Waste Disposal:**

<Description of proposed method of disposal>.

**8. Principal Investigator:**

<Name of Principal Investigator>

.....  
Signature

.....  
Date

\*Categories of Dealings Exempt from Licensing under the Gene Technology Regulations 2001, effective from 1 September 2011

## PART 1 EXEMPT DEALINGS

Item	Description of dealing
1-2	<p>A dealing with a genetically modified <i>Caenorhabditis elegans</i>, unless:</p> <ol style="list-style-type: none"><li>an <i>advantage</i> is conferred on the animal by the genetic modification; or</li><li>as a result of the genetic modification, the animal is capable of secreting or producing an infectious agent.</li></ol>
1-3	<p>A dealing with an animal into which genetically modified somatic cells have been introduced, if:</p> <ol style="list-style-type: none"><li>the somatic cells are not capable of giving rise to infectious agents as a result of the genetic modification; and</li><li>the animal is not infected with a virus that is capable of recombining with the genetically modified nucleic acid in the somatic cells.</li></ol>
1-3A	<p>A dealing with an animal whose somatic cells have been genetically modified <i>in vivo</i> by a replication defective viral vector, if</p> <ol style="list-style-type: none"><li>the <i>in vivo</i> modification occurred as part of a previous dealing; and</li><li>the replication defective viral vector is no longer in the animal; and</li><li>no germ line cells have been genetically modified; and</li><li>the somatic cells cannot give rise to infectious agents as a result of the genetic modification; and</li><li>the animal is not infected with a virus that can recombine with the genetically modified nucleic acid in the somatic cells of the animal</li></ol> <ol style="list-style-type: none"><li>Subject to subitem (2), a dealing involving a host/vector system mentioned in Part 2 of this Schedule and producing no more than 25 litres of GMO culture in each vessel containing the resultant culture.</li><li>The donor nucleic acid:<ol style="list-style-type: none"><li>must meet either of the following requirements:<ol style="list-style-type: none"><li>it must not be derived from organisms implicated in, or with a history of causing, disease in otherwise healthy:<ol style="list-style-type: none"><li>human beings; or</li><li>animals; or</li><li>plants; or</li><li>fungi;</li></ol>it must be characterised and the information derived from its characterisation show that it is unlikely to increase the capacity of the host or vector to cause harm</li></ol></li></ol></li></ol>
1-4	<p><i>Example</i></p> <p>Donor nucleic acid would not comply with subparagraph (ii) if its characterisation shows that, in relation to the capacity of the host or vector to cause harm, it:</p> <ol style="list-style-type: none"><li>provides an advantage; or</li><li>adds a potential host species or mode of transmission; or</li><li>increases its virulence, pathogenicity or transmissibility; and</li><li>must not code for a toxin with an LD<sub>50</sub> of less than 100 microg/kg; and</li><li>must not code for a toxin with an LD<sub>50</sub> of 100 microg/kg or more, if the intention is to express the toxin at high levels; and</li><li>must not be uncharacterised nucleic acid from a toxin-producing organism; and</li><li>must not include a viral sequence, unless the donor nucleic acid:<ol style="list-style-type: none"><li>is missing at least 1 gene essential for viral multiplication that (A) is not available in the cell into which the nucleic acid is introduced; and (B) will not become available during the dealing; and</li><li>cannot restore replication competence to the vector.</li></ol></li></ol>
1-5	<p>A dealing involving shot-gun cloning, or the preparation of a cDNA library, in a host/vector system mentioned in item 1 of Part 2 of this Schedule, if the donor nucleic acid is not derived from either:</p> <ol style="list-style-type: none"><li>a pathogen; or</li><li>a toxin-producing organism</li></ol>

## PART 2 HOST/VECTOR SYSTEMS FOR EXEMPT DEALINGS

Item	Class	Host	Vector
2-1	Bacteria	<p><i>Escherichia coli</i> K12, <i>E. coli</i> B, <i>E. coli</i> C or <i>E. coli</i> Nissle 1917 – any derivative that does not contain:</p> <ol style="list-style-type: none"> <li>generalised transducing phages; or</li> <li>genes able to complement the conjugation defect in a non-conjugative plasmid</li> </ol>	<ol style="list-style-type: none"> <li>Non-conjugative plasmids</li> <li>Bacteriophage               <ol style="list-style-type: none"> <li>lambda</li> <li>lambdoid</li> <li>Fd or F1 (eg M13)</li> </ol> </li> <li>None (non-vector systems)</li> </ol>
		<p><i>Bacillus</i> – specified species – asporogenic strains with a reversion frequency of less than 10<sup>-7</sup>:</p> <ol style="list-style-type: none"> <li><i>B. amyloliquefaciens</i></li> <li><i>B. licheniformis</i></li> <li><i>B. pumilus</i></li> <li><i>B. subtilis</i></li> <li><i>B. thuringiensis</i></li> </ol>	<ol style="list-style-type: none"> <li>Non-conjugative plasmids</li> <li>Plasmids and phages whose host range does not include <i>B. cereus</i>, <i>B. anthracis</i> or any other pathogenic strain of <i>Bacillus</i></li> <li>None (non-vector systems)</li> </ol>
		<i>Pseudomonas putida</i> – strain KT 2440	<ol style="list-style-type: none"> <li>Non-conjugative plasmids including certified plasmids: pKT 262, pKT 263, pKT 264</li> <li>None (non-vector systems)</li> </ol>
		<p><i>Streptomyces</i> – specified species:</p> <ol style="list-style-type: none"> <li><i>S. aureofaciens</i></li> <li><i>S. coelicolor</i></li> <li><i>S. cyaneus</i></li> <li><i>S. griseus</i></li> <li><i>S. lividans</i></li> <li><i>S. parvulus</i></li> <li><i>S. rimosus</i></li> <li><i>S. venezuelae</i></li> </ol>	<ol style="list-style-type: none"> <li>Non-conjugative plasmids</li> <li>Certified plasmids: SCP2, SLP1, SLP2, PIJ101 and derivatives</li> <li>Actinophage phi C31 and derivatives</li> <li>None (non-vector systems)</li> </ol>
		<p><i>Agrobacterium radiobacter</i>  <i>Agrobacterium rhizogenes</i> – disarmed strains  <i>Agrobacterium tumefaciens</i> – disarmed strains</p>	<ol style="list-style-type: none"> <li>Non-tumorigenic disarmed Ti plasmid vectors, or Ri plasmid vectors</li> <li>None (non-vector systems)</li> </ol>
		<p><i>Lactobacillus</i>  <i>Lactococcus lactis</i>  <i>Oenococcus oeni</i> syn.  <i>Leuconostoc oeni</i>  <i>Pediococcus</i>  <i>Photobacterium angustum</i>  <i>Pseudoalteromonas tunicata</i>  <i>Rhizobium</i> (including the genus <i>Allorhizobium</i>)  <i>Sphingopyxis alaskensis</i> syn.  <i>Sphingomonas alaskensis</i>  <i>Streptococcus thermophilus</i>  <i>Synechococcus</i> – specified strains:</p> <ol style="list-style-type: none"> <li>PCC 7002</li> <li>PCC 7942</li> <li>WH 8102</li> </ol>	<ol style="list-style-type: none"> <li>Non-conjugative plasmids</li> <li>None (non-vector systems)</li> </ol>

Item	Class	Host	Vector
		<i>Synechocystis</i> species – strain PCC 6803	
		<i>Vibrio cholerae</i> CVD103-HgR	
		<i>Kluyveromyces lactis</i>	
		<i>Neurospora crassa</i> – laboratory strains	
2-2	Fungi	<i>Pichia pastoris</i>	1. All vectors
		<i>Saccharomyces cerevisiae</i>	2. None (non-vector systems)
		<i>Schizosaccharomyces pombe</i>	
		<i>Trichoderma reesei</i>	
		<i>Yarrowia lipolytica</i>	
2-3	Slime moulds	<i>Dictyostelium</i> species	1. <i>Dictyostelium</i> shuttle vectors, including those based on the endogenous plasmids Ddp1 and Ddp2
			2. None (non-vector systems)
2-4	Tissue culture	Any of the following if they cannot spontaneously generate a whole animal: <ul style="list-style-type: none"> <li>a. animal or human cell cultures (including packaging cell lines);</li> <li>b. isolated cells, isolated tissues or isolated organs, whether animal or human;</li> <li>c. early non-human mammalian embryos cultured <i>in vitro</i></li> </ul>	1. Non-conjugative plasmids
			2. Non-viral vectors, or replication defective viral vectors unable to transduce human cells
			3. Baculovirus ( <i>Autographa californica</i> nuclear polyhedrosis virus), polyhedrin minus
			4. None (non-vector systems)
		Either of the following if they are not intended, and are not likely without human intervention, to vegetatively propagate, flower or regenerate into a whole plant: <ul style="list-style-type: none"> <li>a. plant cell cultures;</li> <li>b. isolated plant tissues or organs</li> </ul>	1. Non-tumorigenic disarmed Ti plasmid vectors, or Ri plasmid vectors, in <i>Agrobacterium tumefaciens</i> , <i>Agrobacterium radiobacter</i> or <i>Agrobacterium rhizogenes</i>
			2. Non-pathogenic viral vectors
			3. None (non-vector systems)